CHEMOENZYMATIC SYNTHESIS OF OPTICALLY ACTIVE α-TOCOPHEROL SIDE CHAIN

Kunihiko Takabe*, Hiroyuki Sawada, Tetsuo Satani, Takashi Yamada, Takao Katagiri, and Hidemi Yoda Department of Applied Chemistry, Faculty of Engineering, Shizuoka University, Hamamatsu 432, JAPAN

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Abstract: Chemoenzymatic synthesis of (2R,6R)-2,6,10-trimethylundecanol and (3R,7R)-3,7,11-trimethyldodecanol, which are key intermediates in the total synthesis of natural α -tocopherol, via Lipase-catalyzed asymmetric induction to prochiral 1,3-propanediol derivatives is described.

(2R,4R',8'R)-α-Tocopherol (1) and closely related substances being potent antioxidants and radical scavengers in biological and chemical systems have been receiving considerable attention with respect to clinical and nutritional applications in human health. In a recent continuation of our work on the synthesis² of 1, we have revealed efficient synthetic methods of some key intermediates of 1, the chroman and the side chain, using asymmetric isomerization^{3a}. Sharpless oxidation^{3b} and asymmetric halolactonization.^{3c}. In the last years many reports⁴ have appeared describing that enzymes are particularly attractive catalysts having the ability to discriminate between two enantiotopic groups of a symmetrical substrate.

With these considerations in mind, this paper describes an efficient chemoenzymatic aproach to the enantioselective synthesis of the α-tocophrol side chains. Our strategy, the route A or B, is based on the Lipase-catalyzed asymmetric induction to the readily accessible prochiral 1,3-propanediol derivatives, 2 and 3.

(R)-6,10-Dimethyl-2-(hydroxymetyl)undecanol (2), which was easily prepared from (R)-citronellal (4) by the modified method reported by Oku et al., 2b was treated with phenylacetate in the presence of Lipase PS⁵ at 15°C

158 K. Takabe et al.

for 3 hours to afford the monoester 5 (86% e.e.)⁶ in 85% yield as shown in Scheme 1. The absolute configuration at C-2 position of 5 was determined to be **R** by its conversion into the known (2S,6R)-2,6,10-trimethylundecanol (6)^{7,8}(85% e.e.⁹). In order to synthesize (2R,6R)-6, the natural α -tocopherol C₁₄-side chain, the enzymatic hydrolysis of the diester 7 was also examined, i.e. porcine pancreatic lipase(PPL)-catalyzed hydrolysis of 7 in phosphate buffer at pH 7.0 gave the monoester 5¹⁰(53% yield, 88% e.e.^{6b}), which was treated with mesyl choride-pyridine, followed by LiAlH4 to afford (2R,6R)-6 (88% e.e.) in 89% yield.¹¹

Moreover, to obtain the optically active C₁₅-side chain of 1, the route B was also investigated as shown in Scheme 2. The diol 9, which was easily prepared by the treatment of diethyl malonate anion with the bromoketal 8, followed by LiAlH4 was subjected to Lipase PS-catalyzed transesterification to give the monoester 10 (88% yield, 98% e.e.). ^{12,13} The absolute configuration of 10 was determined as follows; the monoester 10 was treated with Jones Reagent to give directly (S)-3-acetoxymethylbutanolide (11)^{14,15} in 63% yield. On the basis of this result, the absolute configuration of 10 was identified to be R.

As an useful chiral building block was obtained in hand, the following approach for the conversion of 10 into the C₁₅-side chain was examined. The monoester 10 was treated with mesyl chloride-pyridine, followed by LiAlH4 to afford the alcohol 12¹⁶ (87% yield, 98% e.e. ¹⁷). The treatment of 12 with tosyl chloride-pyridine gave the tosylate 13¹⁸ (85% yield), which was then treated with (R)-3,7-dimethyloctanylmagnesium bromide in the presence of Li₂CuCl₄ ¹⁹ to afford the C-15 ketal 14²⁰ in 85% yield. Deprotection of 14 by hydrochloric acid gave the aldehyde 15²¹ (99% yield), which was reduced by NaBH4 to afford (3R,7R)-3,7,11-trimethyldodecanol (16) in 94% yield.²²⁻²⁴

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Scheme 2.

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- 6. a) (2R,6R)-5: $[\alpha]_D^{20}$ +8.57°(c1.14,CHCl3), b) E.e. was determined by HPLC (Chiralcel OD, hexane-2-propanol=95:5, 0.5 ml/min).
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160 K. TAKABE et al.

- 8. (2S,6R)-6: $[\alpha]_D^{22}$ -7.21°(c1.08,hexane), [Lit.^{7a} $[\alpha]_D^{25}$ -9.21°(c2,hexane)].
- 9. E.e. was determined by HPLC (Chiralcel OD, hexane-ethanol=95:5, 0.2 ml/min).
- 10. (2S,6R)-5: $[\alpha]_D^{22}$ -9.43°(c1.17, CHCl3).
- 11. $(2R,6R)-6:[\alpha]D^{22}+7.67^{\circ}(c1.02,hexane)$ [Lit.[$\alpha]D^{25}+9.02(c0.665,hexane)^{2b},[\alpha]D^{25}+9.13(c2,hexane)^{7a}$]
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- 13. a) $[\alpha]D^{21}+13.1^{\circ}(c1.22,CCl_4)$, b) E.e. was determined by HPLC: Chiralcel OJ, hexane-2-propanol=95:5, 0.7 m1/min.
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- 15. (S)-11: $[\alpha]_D^{22}$ +37.0°(c1.28,CHCl₃) [Lit. ¹⁴ $[\alpha]_D^{24}$ -25.3°(c0.96,CHCl₃) for (R)-11].
- 16. **12**: $[\alpha]D^{24}$ -12.8°(c1.15, CCl4).
- 17. E.e. was determined by HPLC analysis of the (S)-(-)-MTPA ester obtained from 12 (Chirakel OJ, hexane-2-propanol=95:5, 0.2 ml/min).
- 18. 13: $[\alpha]D^{22}+5.0^{\circ}(c1.20,CHC13)$.
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- 20. **14**: $[\alpha]D^{15}+3.04^{\circ}(c1.21, CHCl_3)$,
- 21. **15:** $[\alpha]D^{22}+8.38^{\circ}(c1.37, octane)$ [Lit. $^{3a}[\alpha]D^{25}+8.7^{\circ}(c0.85, octane)],$
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- 23. (3R,7R)-16: $[\alpha]D^{21}+3.26^{\circ}(c1.09, CHCl3)$ [Lit. $[\alpha]D^{18}+3.49^{\circ}(c1.09, CHCl_3)^{22a}, [\alpha]D^{23}+3.35^{\circ}(c0.955, CHCl_3)^{22b}]$.:
- 24. The following conversion of 12 into (4R,7R)-16 was also examined.

d), e), f)
(3R,7R)-16

Y: 42%
[
$$\alpha$$
]_D²⁶ +3.28(c0.67,CHCl₃)

a) $(PhS)_2/Ph_3P$, b) mcpba, c)BuLi, $B_1 \sim \overline{}$, d) Li/EtNH₂

e) HCl, f)NaBH4